A Description of Antimicrobial Susceptibility of Streptococcus pneumoniae-Siriraj Hospital, Thailand: 2008

Somporn Srifuengfung PhD*, Kulkanya Chokephaibulkit MD**, Chanwit Tribuddharat MD, PhD*, Sopita Comerungsee MSc*

* Department of Microbiology, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkok, Thailand ** Department of Pediatrics, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkok, Thailand

Streptococcus pneumoniae was isolated from 170 patient specimens at Siriraj Hospital during January-December 2008. Patients were 66% male and ranged in age from 3 months to 94 years (mean \pm SD = 38.2 \pm 31.7). The largest proportion (29.4%) of isolates were from patients older than 60 years, followed by patients aged 2-5 years (20%) and from patients less than 2 years (12.4%). Monthly isolation was highest in December (22 isolates in December compared to the average of 13 isolates of the other months). Antimicrobial susceptibility for eight drugs was determined by the disk diffusion method. Overall, susceptibility was generally high to chloramphenicol (71.8%), linezolid (100%), ofloxacin (93.5%) and vancomycin (100%), but less susceptible to erythromycin (35.3%), penicillin (31.1%), tetracycline (28.8%) and trimethoprim/ sulfamethoxazole (24.1%). Among the 105 (62%) isolates resistant to three or more drugs, the most common resistance pattern was erythromycin–penicillin–tetracycline–trimethoprim/sulfamethoxazole, accounting for 39% of such isolates, followed by chloramphenicol_erythromycin–penicillin–tetracycline–trimethoprim/sulfamethoxazole (29.5%). The minimal inhibitory concentrations (MIC) of penicillin and cefotaxime were determined by broth microdilution. By 2008 CLSI criteria, 92% and 90% of 51 sterile site isolates were penicillin and cefotaxime susceptible to penicillin and cefotaxime, respectively. The MICs of penicillin were higher for isolates from non-sterile sites than for those from sterile sites.

Keywords: Streptococcus pneumoniae, Antimicrobial susceptibility, Drug resistance

J Med Assoc Thai 2010; 93 (Suppl. 5): S27-S34 Full text. e-Journal: http://www.mat.or.th/journal

Streptococcus pneumoniae is a leading cause of pneumonia, meningitis, otitis media and bacteremia, resulting in substantial morbidity and mortality worldwide⁽¹⁾. In 2005, the World Health Organization estimated that 1.6 million people die of pneumococcal disease every year, including the death of 0.7 to 1 million children aged less than 5 years⁽²⁾. A recent report estimated 11-18 million episodes of serious pneumococcal diseases occurred in the year 2000, causing about 826,000 deaths in children aged less than 5 years⁽³⁾. The emergence of penicillin- and multidrugresistant pneumococcal isolates has been a growing concern. Treatment of penicillin-resistant *S*.

Tribuddharat C,Department of Microbiology, Faculty of Medicine, Siriraj Hospital, Mahidol University, 2 Prannok Rd, Bangkoknoi, Bangkok 10700, Thailand. Phone: 0-2419-9684, Fax: 0-2411-3106 E-mail: sissf@mahidol.ac.th *pneumoniae* has become a challenge, and reports of treatment failure, especially with invasive multidrug-resistant *S. pneumoniae* infections, have increased^(4,5). The aim of this study was to describe the prevalence and the antimicrobial susceptibility patterns of *S. pneumoniae* isolates obtained from patients at Siriraj Hospital in 2008.

Material and Method

S. pneumoniae were isolated from 170 clinical specimens from patients at Siriraj Hospital during January-December 2008. Each isolate represents only one patient. Isolates from the same patient were included only once, giving preference to sterile site isolates. S. pneumoniae isolation was performed by the Bacteriology Laboratory, Department of Microbiology, Faculty of Medicine at Siriraj Hospital, Mahidol University. All isolates were identified as S. pneumoniae based on colony morphology,

Correspondence to:

susceptibility to optochin and bile solubility. They were kept at -70°C in 5% brain heart infusion broth plus 20% glycerol (V/V). Antimicrobial susceptibility testing was performed by plating isolates on Mueller Hinton agar (Becton Dickinson Ltd., USA) supplemented with 5% sheep blood and a nephelometer, and then incubated for 18-20 hours at 35°C in 5% CO, with antimicrobial disks containing chloramphenicol (30 µg), trimethoprim /sulfamethoxazole (1.25/23.75 µg), erythromycin (15 µg), linezolid (10 μ g), ofloxacin (5 μ g), oxacillin (1 μ g), tetracycline (30 μ g) and vancomycin (15 μ g). Susceptibility to each antimicrobial agent was determined using the interpretation criteria for disk diffusion as described by the 2008 Clinical and Laboratory Standards Institute (CLSI) guidelines⁽⁶⁾. S. pneumoniae ATCC 49619 was used for quality control. We defined multi-drug resistance as non-susceptibility to 3 or more classes of antimicrobial agents.

The minimal inhibitory concentrations (MIC) for penicillin and cefotaxime were determined by the broth microdilution method as described by CLSI⁽⁶⁾. Cation-adjusted Mueller Hinton broth (Becton Dickinson Ltd., USA) supplemented with 2-5% lysed horse blood was used. Both drugs were obtained as a standard powder. *S. pneumoniae* ATCC 49619 was used for quality control. Susceptibility results were interpreted using the breakpoints described in the 2008 CLSI guidelines. The comparison between the new⁽⁶⁾ and former⁽⁷⁾ breakpoints is shown in Table 1. The breakpoints for meningitis were applied only to isolates from cerebrospinal fluid (CSF). For isolates from other normally sterile sites, the breakpoints for

"nonmeningitis treated with parenteral penicillin" were used. For isolates from non-sterile sites, breakpoints for "nonmeningitis treated with oral penicillin" were used.

Results

One hundred and seventy isolates of *S. pneumoniae* were collected from clinical specimens of 170 patients (Table 2). Fifty-five isolates were from sterile sites, 46 (83.7%) of which were from blood. Among the 115 isolates from non-sterile sites, 75 (65.2%) were from true sputum of adults with respiratory tract infections. Overall, 66% of isolates were from males; 29.4% were from patients older than 60 years, while 22% were from patients aged less than 2 years and 14.1% were from patients aged 2-5 years (Fig. 1). The highest number of *S. pneumoniae* isolates were obtained in December (Fig. 2). There were 22 isolates in December compared to the average of 13 isolates of the other months.

Based on disk diffusion results (Table 3), all *S. pneumoniae* isolates were susceptible to linezolid and vancomycin; 93.5% were susceptible to ofloxacin while 71.8% were susceptible to chloramphenicol. Less than 50% of isolates were susceptible to erythromycin (35.3%), penicillin (31.1%), tetracycline (28.8%) and trimethoprim/sulfamethoxazole (24.1%). Drug susceptibility of *S. pneumoniae* by specimen sites was shown in Table 4. There is not much difference between sterile and non-sterile sites.

Of 170 isolates, 105 (61.8%) were multi-drug resistant *S. pneumoniae* (Table 5). The most common

Standard	Susceptibility category MIC (µg/ml)			
	Susceptible	Intermediate	Resistant	
Former MIC breakpoints ⁽⁶⁾ from CLSI 2005 (all clinical syndromes, regardless of penicillin administration method) New MIC breakpoints ⁽⁷⁾ from CLSI 2008	≤ 0.06	0.12-1	≥2	
(by clinical syndrome and penicillin route) Meningitis, parenteral penicillin Nonmeningitis, parenteral penicillin Nonmeningitis, oral penicillin	$\leq 0.06 \\ \leq 2 \\ \leq 0.06$	_* 4 0.12-1	$ \geq 0.12 \\ \geq 8 \\ \geq 2 $	

 Table 1. Comparison of former and new MIC breakpoints for determining Streptococcus pneumoniae susceptibility to penicillin, according to CLSI guidelines

MIC = minimal inhibitory concentration, CLSI = Clinical and Laboratory Standards Institute

*No intermediate category for meningitis under new penicillin breakpoints



Fig. 1 Distribution of S. pneumoniae isolates by age group



Fig. 2 Distribution of S. pneumoniae isolates by month

 Table 2. Distribution of S. pneumoniae isolates by specimen sites

Clinical specimens	Number of isolates	%
Sterile sites		
Blood	46	86.8
Cerebrospinal fluid	2	3.8
Plural effusion	3	5.6
Others	2	3.8
Total	53	100
Non-sterile sites		
Sputum	75	64.1
Broncho-alveolar lavage	2	1.7
Nose	9	7.7
Adenoid tissue	7	6.0
Ear	7	6.0
Eye	7	6.0
Sinus	4	3.4
Others	6	5.1
Total	117	100

multi-drug resistance pattern was erythromycinpenicillin-tetracycline-trimethoprim/sulfamethoxazole, accounting for 39% of all multi-drug resistant isolates, followed by chloramphenicol-erythromycinpenicillintetracycline-trimethoprim/sulfamethoxazole (29.5%).

Susceptibilities to penicillin and cefotaxime based on MICs are shown in Table 6. Overall, 92.2% of pneumococcal isolates from sterile sites were susceptible to penicillin; one of 2 CSF isolates and 46 (93.9%) of 49 isolates from other sterile sites were penicillin susceptible. Cefotaxime susceptibility was present in 90.2% of sterile site isolates, including both CSF isolates and 89.8% of isolates from other sterile sites. Among the isolates from non-sterile sites, only 26 (22.6%) had MIC determined. The penicillin and cefotaxime susceptibilities were 26.9% and 76.9%. The MIC₅₀ and MIC₉₀ of penicillin from non-sterile site isolates were higher than that of sterile site isolates.

Discussion

Many clinical studies of pneumococcal disease show a slight male predilection for disease⁽⁸⁾. Although the reasons for this are unclear, our results are consistent with this finding. Children younger than 2 years experience the highest pneumococcal disease burden worldwide. In developed countries, the incidence is highest in those aged 6 months to 1 year. Individuals older than 55-65 years are the second most commonly affected age group worldwide. Immuno compromised persons of any age are at a higher risk for pneumococcal disease⁽⁸⁾. In our study, however, the most common age group from isolates were obtained was > 60 years old. This age distribution of pneumococcal isolation may relate the higher number of elderly compared to pediatric patients seen in our hospital and the fact that sputum, the source of 44% of our isolates, is not routinely collected as part of the evaluation of pediatric patients. However, this study was not necessarily designed to assess differences in disease burden by age, sex, or season, because population denominators are not known and because many of the isolates (non-sterile site) may not have been associated with disease. In addition, we don't get sputum from small children, the number of isolates from adults is certainly going to be higher, regardless of true disease rates. From laboratory culture, as seen in other investigations⁽⁸⁾, S. pneumoniae was isolated in higher frequency in winter.

Antimicrobial resistance in *S. pneumoniae* began to emerge in some parts of the world during the late 1970s⁽⁹⁾. Since then, it has developed into a major public health problem worldwide and raises concern for treatment failures⁽¹⁰⁾. In some settings, *S. pneumoniae* has rapidly developed a clinically relevant resistance to essential antimicrobial agents

Table 3. Drug susceptibility of S. pneumoniae by disk diffusion method

Drugs	Number (%) of susceptibility			
	Susceptible	Intermediate	Resistant	
Chloramphenicol	122 (71.8)	-	48 (28.2)	
Erythromycin	60 (35.3)	-	110 (64.7)	
Linezolid	170 (100)	-	-	
Ofloxacin	159 (93.5)	5 (3.0)	6 (3.5)	
Penicillin (using oxacillin disk)	53 (31.1)	-	117 (68.9)	
Tetracycline	49 (28.8)	8 (4.7)	113 (66.5)	
Trimethoprim/sulfamethoxazole	41 (24.1)	13 (7.7)	116 (68.2)	
Vancomycin	170 (100)	-	-	

Table 4. Drug susceptibility of S. pneumoniae by specimen sites by disk diffusion method

Antimicrobial agents		Percentage of susceptibility ^a				
	Sterile sites $(n = 53)$			Non-sterile sites $(n = 117)$		
	S	Ι	R	S	Ι	R
Chloramphenicol	73.7	-	27.3	70.4	-	29.6
Erythromycin	36.4	-	63.6	35.7	-	64.3
Linezolid	100	-	-	100	-	-
Ofloxacin	98.2	1.8	-	91.3	3.5	5.2
Penicillin (using oxacillin disk)	29.1	-	70.9	32.2	-	67.8
Tetracycline	32.7	1.8	65.5	26.1	6.1	67.8
SXT ^b	30.9	10.9	58.2	20.9	6.1	73
Vancomycin	100	-	-	100	-	-

^aS, susceptible; I, intermediate resistant; R, resistant.

^b Trimethoprim/sulfamethoxazole

(e.g., penicillin, macrolides, third generation cephalosporins, carbapenem drugs and trimetroprim/ sulfamethoxazole)^(5,11,12). A 2004 report from the Asian Network for Surveillance of Resistant Pathogens (ANSORP) showed that about half of S. pneumoniae isolated from normally sterile body sites were nonsusceptible to penicillin⁽¹³⁾. This report included isolates from Thailand in 2000-2001 which showed a high prevalence (53.8%) of penicillin nonsusceptibility (26.9% fully resistant). In the current study, 68.9% of isolates were nonsusceptible to penicillin based on disk diffusion, and all nonsusceptible isolates were fully resistant (Table 3). This 68.9% was higher than the previous value of 63% reported for isolates in Thailand in 2002-2003, but approximately the same to the value of 69% in 2004-2005(14). It is important to point out that by using the disk diffusion method, penicillin resistance occurred in isolates from both sterile and non-sterile sites, almost at the same rate (Table 4). The CLSI does not recommend the disk diffusion method, but rather use the MIC method for isolates from sterile sites.

The prevalence of penicillin nonsusceptibility can vary geographically. Compared to results from a European study during 2001-2003 in *S. pneumoniae* from sterile or nonsterile sites in adults ≥ 16 to > 70years, the prevalence of penicillin resistance among *S. pneumoniae* was higher in the current study than it was in Austria (4.4%), Belgium (11.5%), France (47.6%), Germany (6%), Italy (13%), Portugal (19%) and Switzerland (17.3%), whereas it was similar to Spain (61.9%)⁽¹⁵⁾. Our study found a higher prevalence of erythromycin-resistant *S. pneumoniae* (64.7%) than what was described in a previous Thailand study, which found 49.5% resistance in 2002-2003 and 55% in 2004- $2005^{(13)}$. We found that erythromycin-resistant *S. pneumoniae* was also highly resistant to other antimicrobial agents: penicillin (91.8%), trimethoprim/ sulfam ethoxazole (83.7%), and tetracycline (89.1%).

 Table 5. Multi-drug resistant patterns of S. pneumoniae by disk diffusion method

Pattern of drugs	No. of isolates	%
E-P-TET-SXT	41	39
C-E-P-TET-SXT	31	29.5
E-P-TET	12	11.4
C-E-TET-SXT	4	3.8
E-TET-SXT	4	3.8
C-E-OFX-T-TET-SXT	3	2.9
C-E-P-SXT	3	2.9
C-E-OFX-P-SXT	2	1.9
E-P-SXT	2	1.9
C-E-P-TET	1	1
C-OFX-P-SXT	1	1
C-E-TET	1	1
Total	105	100

C, chloramphenicol; E, erythromycin; OFX, ofloxacin; P, penicillin; TET, tetracycline; SXT, trimethoprim/sulfamethoxazole

As in other investigations⁽¹⁶⁾, erythromycin-resistant *S. pneumoniae* isolates were often resistant to multiple antimicrobial agents (data not shown). Non-susceptibility of *S. pneumoniae* to other amtimicrobial agents, including tetracycline and trimethoprim/sulfamethoxazole, was also found in this study. Our findings are consistent with previous studies of *S. pneumoniae* isolates in Europe showing that trimethoprim/sulfamethoxazole resistance has become common⁽¹⁷⁾. Similar to reports from the USA⁽¹⁸⁾ and China⁽¹⁹⁾, our findings demonstrate that ofloxacin resistance is uncommon. However, increasing and inappropriate use of fluoroquinolones may promote emergence of resistance to quinolones and related agents⁽²⁰⁾.

All *S. pneumoniae* isolates were susceptible to linezolid and vancomycin. Studies suggest that linezolid may be a promising option for the treatment of community-acquired pneumonia due to penicillinresistant *S. pneumoniae*⁽²¹⁾. Both linezolid and vancomycin are recommend to use only after treatment with other antimicrobial agents has failed⁽¹⁹⁾. For pneumococcal meningitis, combination therapy with vancomycin and cefotaxime or ceftriaxone is recommended initially until the results of susceptibility testing are available⁽²²⁾.

In this study, resistance to multiple (three or

 Table 6. Penicillin and cefotaxime susceptibilities of S. pneumoniae isolates from sterile and non-sterile sites by broth microdilution

Drugs	%S ^a	%Iª	$%R^{a}$	$MIC_{50}^{\ b}(\mu g/ml)$	$MIC_{_{90}}{}^{_b}(\mu g/ml)$	MIC Range ($\mu g/ml$)
Sterile sites $(n = 51)$						
Penicillin						
Overall $(n = 51)$	92.2	3.9	3.9	0.12	2	0.03-8
CSF(n=2)	50	-	50	0.06	0.5	0.06-0.5
Other sites $(n = 49)$	93.9	4.1	2	0.12	2	0.03-8
Cefotaxime						
Overall $(n = 51)$	90.2	7.8	2	0.25	2	0.03-4
CSF(n=2)	100	-	-	0.25	0.5	0.25-0.5
Other sites $(n = 49)$	89.8	8.2	2	0.25	2	0.03-4
Non-sterile sites $(n = 26)$						
Penicillin	26.9	7.7	65.4	2	4	0.03-16
Cefotaxime	76.9	19.2	3.8	0.5	2	0.03-4

 ${}^{a}S$ = susceptible, I = intermediate, R = resistant

 ${}^{b}\text{MIC}_{50}$ and MIC_{90} are the minimal inhibitory concentration required to inhibit the growth of 50% and 90% of pneumococci, respectively.

The isolates from CSF were determined by meningitis criteria. The isolates from other sterile sites were determined by nonmeningitis with parenteral penicillin criteria. The isolates from nonsterile sites were determined by nonmeningitis with oral penicillin criteria.

more) antimicrobial agents was common among *S. pneumoniae* isolates. In Canada, multiple resistance was observed in 56 of 1,354 invasive pneumococcal isolates (4.1%). The isolates were resistant to beta-lactams (except for four isolates) and to at least two of the following agents: erythromycin, chloramphenicol, trimethoprim/sulfamethoxazole and ofloxacin⁽²³⁾. These findings highlight the importance of judicious and appropriate antibiotic use as well as the value of effective pneumococcal vaccines, which have been shown to lead to reductions in antimicrobial resistance⁽²⁴⁾.

Penicillin susceptibility of invasive pneumococcal isolates (i.e., isolated from normally sterile sites) was high based on MICs using new CLSI guidelines. (However, penicillin susceptibility of invasive isolates was very low based on disk diffusion). This is in contrast to our 2005 report⁽²⁵⁾ which found 88.2% penicillin nonsusceptibility using old CLSI guidelines. The current study was limited by the lack of clinical information to aid the interpretation of laboratory testing. Blood isolates from patients who may have also had meningitis but who did not have CSF isolates, could be wrongly interpreted as susceptible using the non-meningitis criteria. Although this study included only two CSF isolates, findings support current recommendations that penicillin should not be used to treat meningitis and that cefotaxime is still effective, at least based on in vitro testing.

Acknowledgements

We thank Asthma Foundation of Thailand, the expertise and kindness of the technical staffs at the Bacteriology Laboratory, Department of Microbiology, Faculty of Medicine Siriraj Hospital, Mahidol University, especially Mrs. Siriporn Rakdeekae.

References

- Tortora GJ, Funke BR, Case CL. Microbiology: an introduction. 9th ed. San Francisco, CA: Pearson Education; 2007: 333, 646, 716.
- Pneumococcal conjugate vaccine for childhood immunization-WHO position paper. Wkly Epidemiol Rec 2007; 82: 93-104.
- O'Brien KL, Wolfson LJ, Watt JP, Henkle E, Deloria-Knoll M, McCall N, et al. Burden of disease caused by *Streptococcus pneumoniae* in children younger than 5 years: global estimates. Lancet 2009; 374: 893-902.
- 4. Mera RM, Miller LA, Daniels JJ, Weil JG, White AR. Increasing prevalence of multidrug-resistant

Streptococcus pneumoniae in the United States over a 10-year period: Alexander Project. Diagn Microbiol Infect Dis 2005; 51: 195-200.

- 5. Whitney CG, Farley MM, Hadler J, Harrison LH, Lexau C, Reingold A, et al. Increasing prevalence of multidrug-resistant *Streptococcus pneumoniae* in the United States. N Engl J Med 2000; 343: 1917-24.
- Clinical Laboratory Standards Institute (CLSI). Performance standard for antimicrobial and susceptibility testing: 18th information supplement M100-S18. Wayne, PA: CLSI; 2008.
- Clinical Laboratory Standards Institute (CLSI). Performance standard for antimicrobial and susceptibility testing. 15th information supplement M100-S15. Wayne, PA: CLSI; 2005.
- Muench DF, Rajnik M. Pneumococcal infections: eMedicine Infectious Diseases. Pneumococcal in fections [database on the Internet]. 2008 [cited 2008 May 16]. Available from: www.emedicine. medscape.com
- Felmingham D, Farrell DJ, Reinert RR, Morrissey I. Antibacterial resistance among children with community-acquired respiratory tract infections (PROTEKT 1999-2000). J Infect 2004; 48: 39-55.
- Jacobs MR, Felmingham D, Appelbaum PC, Gruneberg RN. The Alexander Project 1998-2000: susceptibility of pathogens isolated from community-acquired respiratory tract infection to commonly used antimicrobial agents. J Antimicrob Chemother 2003; 52: 229-46.
- McEllistrem MC, Adams JM, Shutt K, Sanza LT, Facklam RR, Whitney CG, et al. Erythromycinnonsusceptible *Streptococcus pneumoniae* in children, 1999-2001. Emerg Infect Dis 2005; 11: 969-72.
- Miller ML, Obert CA, Gao G, Daw NC, Flynn P, Tuomanen E. Cephalosporin-resistant pneumococci and sickle cell disease. Emerg Infect Dis 2005; 11: 1192-6.
- Song JH, Jung SI, Ko KS, Kim NY, Son JS, Chang HH, et al. High prevalence of antimicrobial resistance among clinical *Streptococcus pneumoniae* isolates in Asia (an ANSORP study). Antimicrob Agents Chemother 2004; 48: 2101-7.
- 14. Srifuengfung S, Tribuddharat C, Champreeda P, Daniels J, Chokephaibulkit K, Wongwan N, et al. Antimicrobial susceptibility of *Streptococcus pneumoniae* isolated from patients with respiratory tract infections in Thailand. Southeast Asian J Trop Med Public Health 2008; 39: 461-6.

- Reinert RR, Reinert S, van der LM, Cil MY, Al Lahham A, Appelbaum P. Antimicrobial susceptibility of *Streptococcus pneumoniae* in eight European countries from 2001 to 2003. Antimicrob Agents Chemother 2005; 49: 2903-13.
- Farrell DJ, Jenkins SG, Brown SD, Patel M, Lavin BS, Klugman KP. Emergence and spread of *Streptococcus pneumoniae* with erm(B) and mef(A) resistance. Emerg Infect Dis 2005; 11: 851-8.
- 17. Schmitz FJ, Perdikouli M, Beeck A, Verhoef J, Fluit AC. Resistance to trimethoprim-sulfamethoxazole and modifications in genes coding for dihydrofolate reductase and dihydropteroate synthase in European *Streptococcus pneumoniae* isolates. J Antimicrob Chemother 2001; 48: 935-6.
- Jacobs MR, Good CE, Beall B, Bajaksouzian S, Windau AR, Whitney CG. Changes in serotypes and antimicrobial susceptibility of invasive *Streptococcus pneumoniae* strains in Cleveland: a quarter century of experience. J Clin Microbiol 2008; 46: 982-90.
- 19. Yao KH, Yang YH. *Streptococcus pneumoniae* diseases in Chinese children: past, present and future. Vaccine 2008; 26: 4425-33.
- 20. Richter SS, Heilmann KP, Beekmann SE, Miller NJ, Rice CL, Doern GV. The molecular epidemiology of *Streptococcus pneumoniae* with quinolone

resistance mutations. Clin Infect Dis 2005; 40: 225-35.

- 21. Velissariou IM. Linezolid in children: recent patents and advances. Recent Pat Antiinfect Drug Discov 2007; 2: 73-7.
- 22. Pickering LK, Baker CJ, Long SS, Kimberlin DW. Red Book: 2009 Report of the Committee on Infectious Diseases. 28th ed. Elk Grove Village, IL: American Academy of Pediatrics; 2009: 527.
- 23. Jette LP, Delage G, Ringuette L, Allard R, De Wals P, Lamothe F, et al. Surveillance of invasive *Streptococcus pneumoniae* infection in the province of Quebec, Canada, from 1996 to 1998: serotype distribution, Antimicrobial susceptibility, and clinical characteristics. J Clin Microbiol 2001; 39:733-7.
- 24. Kyaw MH, Lynfield R, Schaffner W, Craig AS, Hadler J, Reingold A, et al. Effect of introduction of the pneumococcal conjugate vaccine on drugresistant *Streptococcus pneumoniae*. N Engl J Med 2006; 354: 1455-63.
- 25. Phongsamart W, Srifeungfung S, Dejsirilert S, Chatsuwan T, Nunthapisud P, Treerauthaweeraphong V, et al. Serotype distribution and antimicrobial susceptibility of *S. pneumoniae* causing invasive disease in Thai children younger than 5 years old, 2000-2005. Vaccine 2007; 25: 1275-80.

การศึกษาความชุกและความไวของเชื้อ Streptococcus pneumoniae ซึ่งแยกได้จากผู้ป่วยใน โรงพยาบาลศิริราชต[่]อยาต้านจุลชีพในปี พ.ศ. 2551

สมพร ศรีเพื่องฟุ้ง, กุลกัญญา โชคไพบูลย์กิจ, ชาญวิทย์ ตรีพุทธรัตน์, โสภิตา คำรังษี

การศึกษา เพื่อหาความซุกของเชื้อ Streptococcus pneumoniae ซึ่งแยกได้จากผู้ป่วยในโรงพยาบาลศิริราช ระหว่างเดือนมกราคม-ธันวาคม พ.ศ. 2551 โดยเชื้อแยกได้จากสิ่งส่งตรวจทั้ง sterile และ nonsterile site พบว่ามีเชื้อ จำนวน 170 สายพันธุ์ ที่แยกได้จากผู้ป่วยซึ่งมีอายุระหว่าง 3 เดือน ถึง 94 ปี (mean ± SD = 38.2 ± 31.7 ปี) เพศชายมากกว่าเพศหญิงสองเท่า โดยแยกเชื้อได้บ่อยที่สุดจากผู้ป่วยที่มีอายุมากกว่า 60 ปี (29.4%) แยก เชื้อได้รองลงมาคือ ผู้ป่วยที่มีอายุ 2-5 ปี (20%) และที่มีอายุน้อยกว่า 2 ปี (12.4%) เดือนธันวาคมจะสามารถแยก เชื้อได้บ่อยที่สุด (22 ราย) โดยเปรียบเทียบกับค่าเฉลี่ยของเดือนอื่น ๆ (13 ราย) ผลการศึกษาความไวของเชื้อ ต่อยาต้านจุลชีพ 8 ชนิด ด้วยวิธี disk diffusion พบว่าเชื้อทั้งหมดมีความไวดีต่อยา chloramphenicol (71.8%) , linezolid (100%), ofloxacin (100%) และ vancomycin (100%) แต่ให้ผลมีความไวดีอย่อยา enythromycin (35.3%), penicillin (31.1%), tetracycline (28.8%) และ trimethoprim/sulfamethoxazole (24.1%) แบบแผนของการดื้อยาต้าน จุลชีพพร้อมกันหลายชนิด ที่พบบ่อยที่สุด คือ enythromycin-penicillin-tetracycline-trimethoprim/sulfamethoxazole (39%) และที่พบรองลงมาคือ chloramphenicol-erythromycin-penicillin-tetracycline-trimethoprim/sulfamethoxazole (39%) และที่พบรองลงมาคือ chloramphenicol-erythromycin-penicillin tetracycline-trimethoprim/sulfamethoxazole (39%) และที่คนบรองลงมาคือ chloramphenicol-erythromycin-penicillin tetracycline-trimethoprim/sulfamethoxazole (39%) และที่คนบรองลงมาคือ chloramphenicol-arythromycin-penicillin tetracycline-trimethoprim/ sulfamethoxazole (29.5%) สำหรับผลการศึกษาค่า MIC ของเชื้อต่อยาต้านจุลชีพ 2 ชนิด คือ penicillin และ cefotaxime ด้วยวิธี broth microdilution นั้นพบว่าเชื้อที่แยกได้จาก sterile site จำนวน 51 สายพันธุ์ มีความไวต่อยา penicillin 92% และมีความไวต่อยา cefotaxime 90% ถ้าเป็นกรณีของผู้ป่วยชนิด meningitis ส่วนเชื้อที่แยกได้จาก nonsterile site จำนวน 26 สายพันธุ์มีความไวต่อยา penicillin 26.9% และ cefotaxime 76.9% สำหรับค่า MIC ของเชื้อที่แยกได้จาก nonsterile site ต่อยาต้านจุลชีพทั้ง 2 ชนิดนี้พบว่าลูงกว่าค่า MIC ของเชื้อที่แยกได้จาก sterile site