# **Special Article**

# Free Radicals in Primary Knee Osteoarthritis

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Free radicals have an important role in the pathogenesis of knee osteoarthritis. Reactive oxygen species (ROS) produced by abnormal chondrocyte metabolism exceeds the physiological buffering capacity and results in oxidative stress. The excessive production of ROS can damage proteins, lipids, nucleic acids, and matrix components. They also serve as important intracellular signaling molecules that amplify the inflammatory response. An understanding of oxidative stress involved in this disease might allow the use of antioxidant therapies in the prevention and/or treatment of knee osteoarthritis.

Keywords: Osteoarthritis, Free radicals, Reactive oxygen species, Chondrocytes

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Osteoarthritis (OA) is the most common form of joint disease that currently lacks effective treatment<sup>(1)</sup>. It is also the leading cause of disability among the elderly. It has been estimated that 68% of Americans (aged over 55 yr) and 35-46% of Thais (aged over 60 yr) have OA<sup>(2,3)</sup>. The economic burden attributed to the joint pain and disability of OA amounts to billions of dollars each year in the USA<sup>(4)</sup>. As the population demographic in the US and Thailand changes to one of a predominantly older generation, the increasing prevalence of OA will become a major public health problem. OA is a disease of a whole organ system including, the joint, the cartilage, the subchondral bone, the synovial capsule and membrane and the periarticular (connective and muscular) tissues. The metabolic and structural changes in the articular cartilage are thought to play a leading role in the initiation and the progression of the disease process. Articular cartilage is a highly specialized and uniquely designed biomaterial that forms the smooth, gliding surface of the diarthrodial joints. It consists mostly of an avascular, aneural and an alymphatic matrix which is synthesized by the sparsely distributed resident cells, the chondrocytes<sup>(5)</sup>. The extracellular matrix is extensively composed of collagens (mainly type II collagen) and proteoglycans (mainly aggrecans) that are responsible for the functional properties of cartilage. The adult articular cartilage is in principle working through the biomechanical properties of its extracellular matrix, and the destruction of the extracellular matrix of articular cartilage is the hallmark of OA. However, the chondrocytes play a decisive role as they are solely responsible for matrix turnover and maintenance. An imbalance between the destruction and synthesis of cartilage is thought to be an essential feature of OA cartilage degeneration<sup>(6,7)</sup>.

As adult articular cartilage is an avascular and, thus, perse hypoxic tissue, the cells must be well adapted to this. The implications of this hypoxic environment are hardly understood on the molecular level. Additionally, the role of changes in oxygen  $(O_2)$ levels during the process of cartilage degeneration seems to be of great interest. Oxygen can also be processed into the so-called reactive oxygen species (ROS). ROS are molecules like hydrogen peroxide,  $(H_2O_2)$ , ions like the hypochlorite ion (OCl<sup>-</sup>), radicals like hydroxyl radical (OH) or the superoxide anion  $(O2^{-})$  which is an ion and a radical at the same time. ROS involved both in intracellular signaling for cell physiology, and in cellular destruction<sup>(8)</sup>. Therefore, this review is intended to give an overview of the role of oxidative stress only in primary knee osteoarthritis (knee OA). In the first part, we explained the reactive oxygen species and oxidative stress. The second part,

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we summarized the current evidences of oxidative stress in knee OA.

### Free Radicals and Oxidative Stress

Free radicals and reactive oxygen species (ROS)

Free radicals can be defined as molecules or molecular fragments with an unpaired electron<sup>(9)</sup>. This unpaired electron usually gives a considerable degree of chemical reactivity to the free radicals. The typical reactions of free radicals are (a) electron donation (from a reducing radical) and electron acceptance (for an oxidizing radical) (b) hydrogen abstraction (c) addition reaction (d) self-annihilation reaction (e) disproportionations

Examples are:

Because of the high reactivity of the unpaired electron in free radical molecules, they rapidly react to adjacent molecules such as DNA, protein, and lipids and cause alterations in their structures. Free radical molecules represent a living state from which oxygenderived species such as superoxide  $(O_2^-)$ , hydrogen peroxide  $(H_2O_2)$ , hydroxyl free radical (OH), lipid peroxides, or related species can be easily generated, both intra- and extra-cellularly. Such agents cause various degrees of toxicity in cells and can lead to either transient or irreversible damage<sup>(8)</sup>.

#### **Physiological roles for ROS**

ROS are produced during normal aerobic cell metabolism, have important physiological roles in maintaining cell redox status, and are required for normal cellular metabolism including facilitating intracellular signaling pathways and the activity of transcription factors such as NF- $\kappa$ B, activator protein 1, C-Myb, p53, and hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) (10-12). In addition, ROS produced by phagocytes also seem to have important physiological roles in priming the immune system<sup>(13,14)</sup>.

#### Cellular Oxidant defense mechanism to free radicals

An antioxidant can be most broadly defined as anything that inhibits an oxidative process. This includes the binding of potentially catalytic iron and copper, which can catalyze oxygen free radical production, in storage proteins. Antioxidants can inhibit oxygen free radical production by four mechanisms<sup>(15)</sup>:

- (i) removing the transition metal catalyst
- (ii) breaking chain reactions
- (iii) reducing reactive species
- (iv) scavenging initiating radicals

They do not act independently of one another but rather function co-operatively. The antioxidant has been introduced as a differentiation between primary and secondary antioxidant defense<sup>(16)</sup>. The former includes the activities of catalase, superoxide dismutase, glutathione peroxidase, and DTdiaphorase, as well as small molecules such as ascorbic acid,  $\alpha$ -tocopherol, GSH,  $\beta$ -carotein and uric acid. The latter includes proteolytic- and lipolytic-enzymes<sup>(16)</sup>, as well as the DNA-repaired systems<sup>(17)</sup>. Antioxidants are substances that compete effectively with other oxidizable substrates even when present in low concentrations, thereby protecting other substrates from the damaging effects of ROS.

#### **Evidence for Oxidative Stress in Knee OA**

Several lines of evidence suggest a role of oxidative stress in the pathogenesis of knee OA. Epidemiologic studies have shown an inverse association between dietary intake of antioxidant and OA progression<sup>(18-20)</sup>. A systematic review of randomized clinical trials by Canter et al (2007) also showed either positive or negative efficacy of both vitamin C and vitamin E in the treatment of knee OA(21). Iron, a catalyst for hydroxyl radical production from hydrogen peroxide is present in both synovial tissue and synovial fluid of knee OA<sup>(22-24)</sup>. Several groups have demonstrated increased oxidative enzyme activity along with decreased antioxidant levels in knee OA sera and synovial fluids<sup>(25-29)</sup>. Because of the highly reactive nature of ROS, it is difficult to directly demonstrate their presence in vivo. It is considerably more practical to measure the 'footprints' of ROS, such as their effects on various lipids, proteins, and nucleic acids. Thus, evidence for oxidative stress or footprints in knee OA has in many cases been generated by approaches that detect oxidant-induced changes to these molecules. Studies of osteoarthritis cartilage and its synovial fluid have demonstrated oxidative damage to proteoglycan<sup>(30,31)</sup>, lipid peroxidation products<sup>(32,33)</sup>, and increased carbonyl groups reflective of oxidation damage to proteins<sup>(34)</sup>. Evidence of oxidative damage to cartilage, extracellular collagen, and intracellular DNA has also been demonstrated.

#### Generation of ROS in knee OA

Free radicals are formed disproportionately in

knee OA by abnormal chondrocytes metabolism<sup>(35-37)</sup>. Nitric oxide (NO) and superoxide anion  $(O_{2})$  are the two main ROS radicals can be produced by chondrocytes. These two highly reactive radicals can further produce the derivative radicals, peroxynitrite (ONOO<sup>-</sup>) and hydrogen peroxide  $(H_2O_2)$ , respectively<sup>(38)</sup>. The first radical, NO radicals, are synthesized by NO synthase (NOS) enzyme. In vitro studies demonstrated that there was an induction of NOS enzyme in chondrocytes culture cells<sup>(39,40)</sup>. The second radical, superoxide anion radicals are produced by the enzyme complex NADPH. Articular chondrocytes can produce superoxide anion by this enzyme system<sup>(41-43)</sup>. In the presence of ferrous iron, hydrogen peroxide and superoxide are converted via the Fenton reaction to highly reactive, aqueous soluble hydroxyl radicals by chondrocyte and cartilage<sup>(44)</sup>. Recently, it was reported that chondrocytes can synthesis the enzyme myeloperoxidase. It was suggested that chondrocytes can produce hypochlorous acid<sup>(45)</sup>. In addition, mechanical compression of chondrocytes can produce reactive oxygen species<sup>(46-49)</sup>.

#### Effect of ROS on chondrocyte DNA

Chondrocyte senescence and cartilage ageing are now considered as an important factor contributing to the development of knee OA. The loss of cells is likely to be of multifactorial origin, with both necrosis and apoptosis being responsible<sup>(50,51)</sup>. Oxidative damage can initiate apoptosis through caspase activation and also may lead to irreversible growth arrest<sup>(52)</sup>. NO has long been considered as the primary inducer of chondrocyte apoptosis mediated by caspase-3 and tyrosine kinase activation<sup>(53,54)</sup>. However, it has become clear that NO by itself cannot initiate apoptosis and that the concomitant production of  $O_2^-$  is required, suggesting the role played by ONOO<sup>-</sup> in this process<sup>(55)</sup>.

Oxygen free radical induced genomic instability, including telomere instability and resulting in replicative senescence and dysfunction in human chondrocytes as demonstrated by Yudoh et al (2005)<sup>(31)</sup>. In tissue cultures of articular cartilage explants, lower antioxidative capacity and stronger staining of nitrotyrosine were observed in the degenerating region of knee OA cartilage as compared with the intact region. During continuous culture of chrondrocytes, telomere length, replicative capacity and GAG production were decreased by treatment with ROS. These effects could be corrected by treatment with an antioxidant agent. Grishko et al (2008) reported mitochondrial DNA damage and poor mitochondrial DNA repair capacity for removing damage caused by oxidative stress in isolated human articular cartilage from knee joint OA patients<sup>(56)</sup>.

#### Effect of ROS on matrix protein synthesis

Exposure of the chondrocytes to H<sub>2</sub>O<sub>2</sub> inhibits proteoglycan and DNA synthesis and depletes intracellular adenosine triphosphate (ATP) as a result of a simultaneous inactivation of glyceraldehydes-3phosphate dehydrogenase<sup>(57,58)</sup>. Exogenous nitric oxide (NO) has suppressive effects on the proteoglycan production. Both S-nitroso-N-acetyl-L, D-penicillamine (SNAP: a donor of NO<sup>-</sup>) and SIN-1(SIN-1,3 morpholinosydnimine: a compound generating both NO and  $O_{2}$ ) are reversible and had an inhibitory effect on glycoaminoglycan synthesis<sup>(59,60)</sup>. Superoxide dismutase reverses SIN-1 inhibited GAG synthesis by primary bovine chondrocytes in a monolayer. Pre-treatment of chondrocyte with SIN-1 or ONOO downregulates aggrecan gene expression, suggesting the involvement of ONOO- in the inhibition of aggrecan synthesis<sup>(61)</sup>.

#### Effect of ROS on cartilage matrix breakdown

ROS may cause damage to all matrix components. Several in vitro studies have reported the degradation of cartilaginous tissue slices by ROSgenerating systems. Damage is believed to be due to direct attack of proteoglycan and collagen molecules by free radicals. Incubation of soluble type I collagen with superoxide anion radicals generated by the xanthine oxidase-hypoxanthine system degrades collagen and prevents the formation of fibrils by this collagen<sup>(62,63)</sup>. OH can degrade collagen and modify the amino acid composition<sup>(64)</sup>. Type I collagen exposure to HOCl fails to degrade collagen but induces the formation of cross-links of an unknown nature<sup>(65)</sup>. HOCl also induces hyaluronic acid cleavage and reduces synovial fluid viscosity(66). Recently, it was suggested that lipid peroxides could play a key role in the structural destabilization of cartilage matrix<sup>(32)</sup>.

#### Conclusion

From these in vitro and *in vivo* studies, we can concluded that in knee OA conditions, ROS such as  $H_2O_2$ , NO,  $O_2^{-1}$ , and NO-derived nitrogen species contribute to cartilage degradation by inhibition matrix synthesis, by directly degrading matrix components and by inducing cell death. Altogether, these finding support the concept of antioxidant therapy to treat knee OA.

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#### References

- 1. Brooks PM. Impact of osteoarthritis on individuals and society: how much disability? Social consequences and health economic implications. Curr Opin Rheumatol 2002; 14: 573-7.
- 2. Elders MJ. The increasing impact of arthritis on public health. J Rheumatol Suppl 2000; 60: 6-8.
- Kuptniratsaikul V, Tosayanonda O, Nilganuwong S, Thamalikitkul V. The epidemiology of osteoarthritis of the knee in elderly patients living an urban area of Bangkok. J Med Assoc Thai 2002; 85: 154-61.
- Centers for Disease Control and Prevention (CDC). Update: direct and indirect costs of arthritis and other rheumatic conditions-United States, 1997. MMWR Morb Mortal Wkly Rep 2004; 53: 388-9.
- Muir H. The chondrocyte, architect of cartilage. Biomechanics, structure, function and molecular biology of cartilage matrix macromolecules. Bioessays 1995; 17: 1039-48.
- Aigner T, Vornehm SI, Zeiler G, Dudhia J, von der Mark K, Bayliss MT. Suppression of cartilage matrix gene expression in upper zone chondrocytes of osteoarthritic cartilage. Arthritis Rheum 1997; 40: 562-9.
- Aigner T, Zien A, Gehrsitz A, Gebhard PM, McKenna L. Anabolic and catabolic gene expression pattern analysis in normal versus osteoarthritic cartilage using complementary DNA-array technology. Arthritis Rheum 2001; 44: 2777-89.
- 8. Halliwell B. Reactive oxygen species in living systems: source, biochemistry, and role in human disease. Am J Med 1991; 91: 14S-22S.
- Halliwell B, Gutteridge J. Oxygen is poisonous-an introduction to oxygen toxicdity and free radicals. In: Halliwell B, editor. Free radicals in biology and medicine. 2<sup>nd</sup> ed. Oxford: Oxford University Press; 1989: 1-20.
- Hentze MW, Rouault TA, Harford JB, Klausner RD. Oxidation-reduction and the molecular mechanism of a regulatory RNA-protein interaction. Science 1989; 244: 357-9.
- Groulx I, Lee S. Oxygen-dependent ubiquitination and degradation of hypoxia-inducible factor requires nuclear-cytoplasmic trafficking of the von Hippel-Lindau tumor suppressor protein.

Mol Cell Biol 2002; 22: 5319-36.

- 12. Haddad JJ. Antioxidant and prooxidant mechanisms in the regulation of redox(y)-sensitive transcription factors. Cell Signal 2002; 14: 879-97.
- Babior BM. Oxygen-dependent microbial killing by phagocytes (second of two parts). N Engl J Med 1978; 298: 721-5.
- Babior BM. Oxygen-dependent microbial killing by phagocytes (first of two parts). N Engl J Med 1978;298:659-68.
- Crichton R. Inorganic biochemistry of iron metabolism. Chichester: Ellis Horwood Limited; 1992.
- 16. Cadenas E. Biochemistry of oxygen toxicity. Annu Rev Biochem 1989; 58: 79-110.
- Demple B, Halbrook J. Inducible repair of oxidative DNA damage in Escherichia coli. Nature 1983; 304: 466-8.
- McAlindon TE, Jacques P, Zhang Y, Hannan MT, Aliabadi P, Weissman B, et al. Do antioxidant micronutrients protect against the development and progression of knee osteoarthritis? Arthritis Rheum 1996; 39: 648-56.
- 19. Sowers M. Epidemiology of risk factors for osteoarthritis: systemic factors. Curr Opin Rheumatol 2001; 13: 447-51.
- 20. Zhang Y, Jordan JM. Epidemiology of osteoarthritis. Rheum Dis Clin North Am 2008; 34: 515-29.
- 21. Canter PH, Wider B, Ernst E. The antioxidant vitamins A, C, E and selenium in the treatment of arthritis: a systematic review of randomized clinical trials. Rheumatology (Oxford) 2007; 46: 1223-33.
- 22. Fritz P, Saal JG, Wicherek C, Konig A, Laschner W, Rautenstrauch H. Quantitative photometrical assessment of iron deposits in synovial membranes in different joint diseases. Rheumatol Int 1996; 15: 211-6.
- 23. Yazar M, Sarban S, Kocyigit A, Isikan UE. Synovial fluid and plasma selenium, copper, zinc, and iron concentrations in patients with rheumatoid arthritis and osteoarthritis. Biol Trace Elem Res 2005; 106: 123-32.
- 24. Morris CJ, Blake DR, Wainwright AC, Steven MM. Relationship between iron deposits and tissue damage in the synovium: an ultrastructural study. Ann Rheum Dis 1986; 45: 21-6.
- 25. Sarban S, Kocyigit A, Yazar M, Isikan UE. Plasma total antioxidant capacity, lipid peroxidation, and erythrocyte antioxidant enzyme activities in patients with rheumatoid arthritis and osteoarthritis. Clin

Biochem 2005; 38: 981-6.

- Surapaneni KM, Venkataramana G. Status of lipid peroxidation, glutathione, ascorbic acid, vitamin E and antioxidant enzymes in patients with osteoarthritis. Indian J Med Sci 2007; 61: 9-14.
- 27. Altindag O, Erel O, Aksoy N, Selek S, Celik H, Karaoglanoglu M. Increased oxidative stress and its relation with collagen metabolism in knee osteoarthritis. Rheumatol Int 2007; 27: 339-44.
- Ostalowska A, Birkner E, Wiecha M, Kasperczyk S, Kasperczyk A, Kapolka D, et al. Lipid peroxidation and antioxidant enzymes in synovial fluid of patients with primary and secondary osteoarthritis of the knee joint. Osteoarthritis Cartilage 2006; 14: 139-45.
- 29. Regan EA, Bowler RP, Crapo JD. Joint fluid antioxidants are decreased in osteoarthritic joints compared to joints with macroscopically intact cartilage and subacute injury. Osteoarthritis Cartilage 2008; 16: 515-21.
- 30. Rees MD, Hawkins CL, Davies MJ. Hypochlorite and superoxide radicals can act synergistically to induce fragmentation of hyaluronan and chondroitin sulphates. Biochem J 2004; 381: 175-84.
- 31. Yudoh K, Nguyen T, Nakamura H, Hongo-Masuko K, Kato T, Nishioka K. Potential involvement of oxidative stress in cartilage senescence and development of osteoarthritis: oxidative stress induces chondrocyte telomere instability and downregulation of chondrocyte function. Arthritis Res Ther 2005; 7: R380-91.
- 32. Tiku ML, Shah R, Allison GT. Evidence linking chondrocyte lipid peroxidation to cartilage matrix protein degradation. Possible role in cartilage aging and the pathogenesis of osteoarthritis. J Biol Chem 2000; 275: 20069-76.
- Shah R, Raska K Jr, Tiku ML. The presence of molecular markers of in vivo lipid peroxidation in osteoarthritic cartilage: a pathogenic role in osteoarthritis. Arthritis Rheum 2005; 52: 2799-807.
- Dalle-Donne I, Rossi R, Giustarini D, Milzani A, Colombo R. Protein carbonyl groups as biomarkers of oxidative stress. Clin Chim Acta 2003; 329: 23-38.
- 35. Mazzetti I, Grigolo B, Pulsatelli L, Dolzani P, Silvestri T, Roseti L, et al. Differential roles of nitric oxide and oxygen radicals in chondrocytes affected by osteoarthritis and rheumatoid arthritis. Clin Sci (Lond) 2001; 101: 593-9.
- 36. Henrotin YE, Bruckner P, Pujol JP. The role of reactive oxygen species in homeostasis and degradation of cartilage. Osteoarthritis Cartilage

2003; 11: 747-55.

- Henrotin Y, Kurz B, Aigner T. Oxygen and reactive oxygen species in cartilage degradation: friends or foes? Osteoarthritis Cartilage 2005; 13: 643-54.
- Tiku ML, Liesch JB, Robertson FM. Production of hydrogen peroxide by rabbit articular chondrocytes. Enhancement by cytokines. J Immunol 1990; 145: 690-6.
- Stadler J, Stefanovic-Racic M, Billiar TR, Curran RD, McIntyre LA, Georgescu HI, et al. Articular chondrocytes synthesize nitric oxide in response to cytokines and lipopolysaccharide. J Immunol 1991; 147: 3915-20.
- Palmer RM, Hickery MS, Charles IG, Moncada S, Bayliss MT. Induction of nitric oxide synthase in human chondrocytes. Biochem Biophys Res Commun 1993; 193: 398-405.
- Hiran TS, Moulton PJ, Hancock JT. Detection of superoxide and NADPH oxidase in porcine articular chondrocytes. Free Radic Biol Med 1997; 23:736-43.
- 42. Moulton PJ, Hiran TS, Goldring MB, Hancock JT. Detection of protein and mRNA of various components of the NADPH oxidase complex in an immortalized human chondrocyte line. Br J Rheumatol 1997; 36: 522-9.
- Hiran TS, Moulton PJ, Hancock JT. In situ detection of superoxide anions within porcine articular cartilage. Br J Biomed Sci 1998; 55: 199-203.
- 44. Tiku ML, Yan YP, Chen KY. Hydroxyl radical formation in chondrocytes and cartilage as detected by electron paramagnetic resonance spectroscopy using spin trapping reagents. Free Radic Res 1998; 29: 177-87.
- 45. Attur MG, Dave M, Akamatsu M, Katoh M, Amin AR. Osteoarthritis or osteoarthrosis: the definition of inflammation becomes a semantic issue in the genomic era of molecular medicine. Osteoarthritis Cartilage 2002; 10: 1-4.
- Beecher BR, Martin JA, Pedersen DR, Heiner AD, Buckwalter JA. Antioxidants block cyclic loading induced chondrocyte death. Iowa Orthop J 2007; 27:1-8.
- 47. Kurz B, Lemke A, Kehn M, Domm C, Patwari P, Frank EH, et al. Influence of tissue maturation and antioxidants on the apoptotic response of articular cartilage after injurious compression. Arthritis Rheum 2004; 50: 123-30.
- 48. Healy ZR, Lee NH, Gao X, Goldring MB, Talalay P, Kensler TW, et al. Divergent responses of chondrocytes and endothelial cells to shear

stress: cross-talk among COX-2, the phase 2 response, and apoptosis. Proc Natl Acad Sci U S A 2005; 102: 14010-5.

- Tomiyama T, Fukuda K, Yamazaki K, Hashimoto K, Ueda H, Mori S, et al. Cyclic compression loaded on cartilage explants enhances the production of reactive oxygen species. J Rheumatol 2007; 34: 556-62.
- Blanco FJ, Guitian R, Vazquez-Martul E, de Toro FJ, Galdo F. Osteoarthritis chondrocytes die by apoptosis. A possible pathway for osteoarthritis pathology. Arthritis Rheum 1998; 41: 284-9.
- Horton WE Jr, Feng L, Adams C. Chondrocyte apoptosis in development, aging and disease. Matrix Biol 1998; 17: 107-15.
- 52. Oh M, Fukuda K, Asada S, Yasuda Y, Tanaka S. Concurrent generation of nitric oxide and superoxide inhibits proteoglycan synthesis in bovine articular chondrocytes: involvement of peroxynitrite. J Rheumatol 1998; 25: 2169-74.
- 53. Blanco FJ, Ochs RL, Schwarz H, Lotz M. Chondrocyte apoptosis induced by nitric oxide. Am J Pathol 1995; 146: 75-85.
- 54. Wu GJ, Chen TG, Chang HC, Chiu WT, Chang CC, Chen RM. Nitric oxide from both exogenous and endogenous sources activates mitochondriadependent events and induces insults to human chondrocytes. J Cell Biochem 2007; 101: 1520-31.
- 55. Del Carlo M Jr, Loeser RF. Nitric oxide-mediated chondrocyte cell death requires the generation of additional reactive oxygen species. Arthritis Rheum 2002; 46: 394-403.
- 56. Grishko VI, Ho R, Wilson GL, Pearsall AW. Diminished mitochondrial DNA integrity and repair capacity in OA chondrocytes. Osteoarthritis Cartilage 2009; 17: 107-13.
- 57. Vincent F, Brun H, Clain E, Ronot X, Adolphe M. Effects of oxygen-free radicals on proliferation kinetics of cultured rabbit articular chondrocytes. J Cell Physiol 1989; 141: 262-6.
- 58. Baker MS, Feigan J, Lowther DA. The mechanism

of chondrocyte hydrogen peroxide damage. Depletion of intracellular ATP due to suppression of glycolysis caused by oxidation of glyceraldehyde-3-phosphate dehydrogenase. J Rheumatol 1989; 16: 7-14.

- Taskiran D, Stefanovic-Racic M, Georgescu H, Evans C. Nitric oxide mediates suppression of cartilage proteoglycan synthesis by interleukin-1. Biochem Biophys Res Commun 1994; 200: 142-8.
- 60. Stefanovic-Racic M, Meyers K, Meschter C, Coffey JW, Hoffman RA, Evans CH. Comparison of the nitric oxide synthase inhibitors methylarginine and aminoguanidine as prophylactic and therapeutic agents in rat adjuvant arthritis. J Rheumatol 1995; 22: 1922-8.
- 61. Mathy-Hartert M, Martin G, Devel P, Deby-Dupont G, Pujol JP, Reginster JY, et al. Reactive oxygen species downregulate the expression of pro-inflammatory genes by human chondrocytes. Inflamm Res 2003; 52: 111-8.
- Monboisse JC, Braquet P, Randoux A, Borel JP. Non-enzymatic degradation of acid-soluble calf skin collagen by superoxide ion: protective effect of flavonoids. Biochem Pharmacol 1983; 32: 53-8.
- 63. Bates EJ, Harper GS, Lowther DA, Preston BN. Effect of oxygen-derived reactive species on cartilage proteoglycan-hyaluronate aggregates. Biochem Int 1984; 8: 629-37.
- Monboisse JC, Borel JP. Oxidative damage of collagen. In: Emerit I, Chance B, editors. Free radicals and aging. Basel: Birkhauser Verlag; 1992: 323-8.
- 65. Green SP, Baker MS, Lowther DA. Depolymerization of synovial fluid hyaluronic acid (HA) by the complete myeloperoxidase (MPO) system may involve the formation of a HA-MPO ionic complex. J Rheumatol 1990; 17: 1670-5.
- Saari H, Konttinen YT, Friman C, Sorsa T. Differential effects of reactive oxygen species on native synovial fluid and purified human umbilical cord hyaluronate. Inflammation 1993; 17: 403-15.

# อนุมูลอิสระในโรคข้อเข่าเสื่อมปฐมภูมิ

# วีระศักดิ์ สุทธิพรพลางกูร, นพวรรณ ภู่มาลา มอราเลส, ทศศาสตร์ หาญรุ่งโรจน์

อนุมูลอิสระมีบทบาทสำคัญในกลไกการเกิดโรคข้อเข่าเสื่อม อนุมูลอิสระของออกซิเจนที่ผลิตจาก เมตาบอลิสมที่ผิดปกติของเซลล์กระดูกอ่อนที่เกินจากภาวะปกติของร่างกายทำให้เกิดภาวะเครียดออกซิเดชั่น อนุมูลอิสระที่มากเกินปกติสามารถทำลายทั้งโปรตีน, ไขมัน, ดีเอนเอ และส่วนประกอบของผิวข้อ นอกจากนี้ อนุมูลอิสระยังส่งเสริมให้เกิดการอับเสบที่มากขึ้นโดยผ่านการสื่อสารภายในเซลล์ ดังนั้นการเข้าใจในภาวะ เครียดออกซิเดชั่น ในโรคข้อเข่าเสื่อมอาจนำไปสู่การให้การป้องกัน หรือ รักษาโดยใช้สารต้านภาวะเครียดออกซิเดชั่น ต่อไป